

PATENT

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Field of the invention

The present invention is also related to an identification and/or quantification apparatus based upon said method, that allows the identification and/or the quantification of positive locations of bounded target compounds upon said chips.

Biological assays are mainly based upon interaction specificity between two biological molecules such two strands of nucleic acid molecules, an antigen and an antibody or a ligand and its receptor. The present challenge of biological assays is to perform simultaneously the multiple detection of molecules present in a sample. Miniaturisation and development of arrays upon the surface of "biochips" are tools that allow multiplex reactions in a microscopic format, said detection being made with a limited volume of sample for the screening and/or the

identification of multiple possible target compounds. These arrays are formed of discrete regions, containing a specific capture molecule used for the binding of the target compound. These discrete regions, as small as a few
5 micrometers, allow the fixation of several thousands capture molecules per cm² surface (WO 95/11995).

However, the detection of bounded target compounds is difficult, since their amount is very small due to said miniaturisation (few femtomoles or even few
10 attomoles). Therefore, only extremely sensitive methods are adequate for such detection.

It has been proposed a labelling of a target compound like DNA with fluorescent molecules after their possible genetic amplification. When an RNA molecule has to
15 be detected, it is first transformed into a cDNA, before its possible amplification. If direct labelling of the target compound is not possible, a double reaction (sandwich reaction) can be performed. However, the amount of fluorescent molecules is so low that it is necessary to
20 develop specific array scanners for the detection and/or the quantification of the bounded compound upon the "hybridisation chips". Said expensive specific scanners comprise a laser scanner for excitation of the fluorescent molecules, a pinhole for decreasing the noise fluorescent
25 background, and a photomultiplier for increasing the sensitivity of the detection.

Alternative detection methods that present a high sensitivity are described in the documents US-5,821,060 and WO95/04160 and are based upon the
30 detection using expensive devices such as mass spectrometers.

It has also been proposed methods based upon the precipitation of specific products resulting of a

colorimetric labelling (US 5,270,167, US 4,731,325, EP-A-0301141) or the result of an enzymatic activity (EP-A-0393868, WO 86/02733, EP-A-0063810). However, said methods are either characterized by a low sensitivity or
5 are not adequate for the detection of a target compound upon "hybridisation chips", because the precipitate will occur at a certain distance of the reaction binding and its location can not be easily correlated with a specific bounded target compound. In addition, the density of the
10 precipitate of such enzymatic reactions is not enough opaque for allowing a detection by light absorption.

It has also been proposed to improve the detection by fixing a soluble product obtained from the enzymatic reaction with a metal before its precipitation.
15 However, as the result of said enzymatic reaction is a soluble product, there is no correlation between the location of the precipitate and the detection of a specific bounded target compound.

20 Aims of the invention

The present invention aims to provide a new identification and/or quantification method of one or more target compounds present (possibly simultaneously) in a biological sample and that will not present the drawbacks
25 of the state of the art.

The present invention aims to provide such a method that is simple and not expensive, that allows the detection of said target compounds by using fixed capture molecules upon arrays of the surface of a solid support.

30 A last aim of the present invention is to provide also a simple and non-expensive apparatus based upon said method, that improves the identification and/or the quantification of bounded target compounds upon "hybridisation chips".

made of glasses, filters, electronic device, polymeric or metallic materials, etc. Preferably, said arrays contain specific locations (advantageously presented according to a specific pattern), each of them containing normally only one species of capture molecule.

The fixation of DNA strands on proteins thereafter specifically attached to sites specific locations on a substrate, is described in the document US-5,561,071. It is also known that capture chemicals can be linked to microtubes that are then spatially arranged in order to produce an array, as described in the document GB-3 319 838, or to obtain the direct synthesis of oligonucleotides on specific surfaces by using photolithographic techniques as described in the documents EP-0476014, US-5,510,270, US-5,445,934, WO97/29212, US-5,605,662, US-5,632,957 and WO94/22889.

All these methods for the fixation of capture molecules on the surface of a solid support in order to obtain the above-described arrays are compatible with the present invention.

The biological target compounds according to the invention may be present in a biological (or possibly a non-biological) sample such as clinical samples extracted from blood, urine, faeces, saliva, pus, serum, tissues, fermentation solutions or culture media. Said target compounds are preferably isolated purified, cleaved, copied and/or amplified if necessary by known methods by the person skilled in the art before their detection and/or quantification upon the "hybridisation chips".

Preferably, the formation of a precipitate at the location of the binding is obtained with the fixation of a metallic compound upon the bounded target compound or by the result of a reduction of a metal in the presence of an enzyme. Advantageously, a reduction of silver in the

presence of colloidal gold allows the formation of a precipitate at a distance not exceeding few micrometers from the bounded target compound to its capture molecule.

According to the invention, the specific locations on the array are smaller than 1000 μm in length. These locations or spots have preferably a diameter comprised between 10 and 500 μm and are separated by distance of similar order of magnitude, so that the array of the solid support comprises between 100 and 250,000 spots upon the surface of 1 cm^2 . However, it is also possible to prepare spots smaller as 1 μm or less upon which the capture molecules are fixed. The formation of said spots or locations would be obtained by known microelectronic or photolithographic processes and devices that allow the fixation of said capture molecules on the surface of the solid support either by a covalent linkage or a non-covalent adsorption. The covalent linkage technique is preferred in order to control specifically the sites of capture molecules fixation and avoid possible drawbacks that may result with several capture molecules (like nucleic acids or antibodies) that can be desorbed during incubation or washing step.

One of the preferred embodiment is the fixation of biological molecules like proteins, peptides or nucleic acid sequences by linkage of amino groups on activated glass bearing aldehyde moiety. The incorporation of an amine group in the nucleic acid chain is easily obtained using aminated nucleotide during their synthesis. Aminated amino acids can be fixed upon the surface of a solid support like glass bearing aldehyde groups as described by Schena et al. (Proc. Natl. Acad. Sci. USA, 93, pp. 10614-10619 (1996)) or as described in the document US-5,605,662 and the publication of Krensky et al. (Nucleic

Acids Research, 15, pp. 2891-2909 (1987)). The linkage between an amino and a carboxyl group is obtained by the presence of a coupling agent like carbodiimide compounds as described by Joos et al. (Anal. Biochem., 247, pp. 96-101 (1997)). Thiol modified oligonucleotides can be used also to obtain a reaction with amino groups upon the surface of a solid support in the presence of cross-linking molecules (Thrisey et al., Nucleic Acids Research, 24, pp. 3031-3039 (1996)). Similarly, oligonucleotides can be fixed to a gel like polyacrylamide bearing hydroxyl and aldehyde groups as described in the document US-5,552,270 and WO98/28444.

The binding (or recognition) of the target compound upon their corresponding specific capture molecules is a spontaneous non-covalent reaction when performed in optimal conditions. It involves non-covalent chemical bindings. The medium composition and other physical and chemical factors affect the rate and the strength of the binding. For example for nucleotide strand recognition, low stringency and high temperature lower the rate and the strength of the binding between the two complementary strands. However, they also very much lower the non specific binding between two strands (which are not perfectly complementary). When several sequences are similar, the specificity of the binding can be enhanced by addition of a small amount of non-labelled molecules, which will compete with their complementary sequence, but much more with the other ones, thus lowering the level of cross-reactions.

The optimisation of the binding conditions is also necessary for antigen/antibody or ligand/receptors recognition, but they are usually rather specific.

A preferred embodiment of this invention is to take party of the amplification given by the catalytic

reduction of Ag^+ in the contact of other metals like gold. Gold nanoparticles are currently available and they can be easily fixed to molecules like protein. For example, streptavidin coated gold particles are available on the
5 market.

According to a preferred embodiment of this invention, one uses a labelled target molecule, which is then recognised by a conjugate. This labelled molecule (biotin, haptens, ...) can be considered as a first member
10 of the binding pair. For DNA, the labelling is easily done by incorporation of biotinylated nucleotides during their amplification. For the RNA, biotinylated nucleotides are used for their copy in cDNA or thereafter in the amplification step. Amplification of the nucleotide
15 sequences is a common practice since the target molecules are often present in very low concentrations. Proteins are easily labelled using NHS-biotin or other reactions. Once the biotinylated molecules are captured, a streptavidin-gold complex, which is the second member of the binding
20 pair, is added and the streptavidin specifically recognises biotin, so that the complex is fixed at the location where the target is fixed. If haptens are used as label, an antibody-gold complex will be used. Then a reactive mixture containing Ag^+ and a reducing agent is added on the surface
25 and Ag layers will precipitate on the gold particles leading to the formation of crystal particles.

Direct labelling of the target molecules with gold is possible by using gold-labelled antigens, antibodies or nucleotides.

30 An alternative is to avoid any labelling of the target molecule, and then a second nucleotide sequence is used which is labelled. They then formed a sandwich hybridisation or a sandwich reaction with the capture

molecule fixing the target and the labelled nucleotide sequence, which allows the detection to go on. Like above, the labelled nucleotide sequence is able to catalyse itself the precipitation of the metal or it does it through a second complex.

The Ag precipitation corresponds to the location of the binding of biotinylated nucleotide sequence. As said location is well defined, it is possible to identify the presence of said precipitate (specific spot of the array).

The precipitate has the form of small crystals that reach with time a diameter of about 1 μ m. The formation of these small crystals represents a real amplification of the signal since they originated from the presence of gold particles a few nm in diameter.

Unexpectedly, within a given range of labelled nucleotide sequences present on the surface, a concentration curve could be obtained between the gold-labelled nucleotide sequence concentration and the amount of precipitate on the surface. One constraint of the array is that the detection signal has to be correlated with the location where it originates.

Because of its granular form, the precipitate advantageously modifies the reflection of the light. It also leads to a strong diffusion of the light (spot detected), which is recordable by known detection means. Such diffusion assays are typically detected and recorded from the reflection of a light beam with photodiodes. One unexpected observation is that this assay for the presence of silver crystals was found unexpectedly very sensitive. The fact that the silver precipitate appears as a black surface allows the use of a scanner (absorption of the light through the transparent surface of the array). The presence of insoluble precipitate will absorb the light,

which is then detected and recorded. The advantage of the scanner is that only a small portion of the array is detected at one time so that a much better resolution can be obtained. Either the illumination beam or the detection
5 surface is focalised and the signal is recorded so that the image of the array can be reconstituted. The detection means (detector) can be a CCD or CMOS camera, which measures the overall array. The resolution of the detection is then dependent on the number of pixels of the camera. On
10 the other end, the detector can be constituted of photo-diodes arranged into a line and the image is scanned by moving in front of this line. Scanners with a sensitivity of 11 μm for a pixel can be constructed, which are sufficient to analyse spots of 50 μm in diameter or bigger.

15 A full illumination of the array combined with a recording of the light transmitted is also possible (faster than the scanner, but seems to be less sensitive).

As a metal, silver is able to reflect light by itself. Even if the efficiency of this reflection is
20 low, it exists and can be used in order to localise the silver precipitate (spots). Because of its metal nature, other methods like variations of an electromagnetic field or electric conductance are also possible.

Another aspect of the present invention
25 concerns a diagnostic and/or quantification apparatus of one or more identical or different target compound(s) obtained from a sample, said apparatus comprising:

- a detection and/or quantification device of precipitates (spots) upon the surface of a solid support resulting
30 from the binding of a target compound upon its corresponding capture molecule above-described,
- possibly a reading device of information(s) recorded upon said solid support (such as barcodes), and

- a computer programmed to:
- possibly recognise the discrete regions bearing capture molecules,
- collect the results obtained from said detection
- 5 device, possibly correlated with the information(s) obtained from said reading device, and
- carry out a diagnostic and/or quantification of said target compound(s).

Hence, detection resolution, and more

10 particularly the reliability of the final quantification depends largely on the characteristics of the detection device. Especially, when the detection device includes a CCD camera, the reliability depends on its number of pixels. The number of pixels thus limits the allowed

15 sensitivity of the quantification. Typically, it is possible to obtain with a CCD a resolution of 10 μm for a pixel, which are sufficient to analyse spots of 100 μm in diameter or bigger. However, such quantification is limited by the number of pixels, by the resolution of each pixel

20 and the fact that the sensitivity is given by only one view point. One view point depends on the three following patterns : the position of the lecture element like CCD camera, the position of the object to be detected and the position of the lightening of the object.

25 In order to respond to said objectives, the present invention is also related to a method (preferably dedicated to the detection and/or the quantification of a precipitate according to the invention, but not only to such precipitate) for the quantification of a volume of a

30 precipitate (preferably containing metal crystals) upon a defined surface of a solid support, said defined surface of a solid support being defined by an array of at least 4, at least 10, at least 16, at least 20 or more discrete regions

per cm², each discrete region possibly comprising a precipitate. According to the invention images of said defined surface comprising one or more precipitates correspond to different views, said images containing
 5 analogue informations being taken by one or several camera(s) and upon illumination by one or several illuminant source(s) being spatially arranged relatively to each other according to a predetermined pattern; the corresponding image analogue informations of said defined
 10 surface comprising said precipitate(s) being transformed and converted into digital form or set of digital forms and compared to a first and to a second reference standard to determine the volume of the precipitate(s) to be quantified.

15 The first reference standard corresponds to the digital form or set of digital forms obtained from analogue informations contained in images taken on said surface without precipitate.

20 The second reference standard corresponds to the digital form or set of digital forms obtained from analogue informations contained in images taken on said surface comprising precipitates of known volume.

The term "volume" should be understood to mean the volume for which it is desired to obtain
 25 dimensional-type information. In the present invention, said volume results from a chemical or biochemical reaction following a binding between a target compound and its corresponding compound. Therefore, said obtained volume is the expression of said chemical or biochemical reaction
 30 following a binding between a target compound and its corresponding capture compound.

The term "image" should be understood to mean a group of pixels which is an illustration of a measure of

said volume and which may be directly transmitted to and registered upon a monitor such as a screen or a printer.

The present invention is also related to an apparatus comprising means for implementing said method, preferably comprising one or several sensor(s) provided with cameras and with one and/or several illuminant source(s) which are spatially arranged relatively to each other according to a predetermined pattern and which are associated with an analogue information acquisition system, the information being measured by using said sensors and being converted into digital form by a processing unit.

Preferably the transformation and conversion are made by a processing unit on board of the camera or in a computer.

15 The cameras are preferably mono, infrared, colour, special adjacent range CCD or CMOS cameras or similar lecture technologies.

The illuminant source is preferably an infrared light having a wavelength similar to the dimension of the metal crystals contained in the precipitate(s) and which is advantageously produced by using a single diode or diodes having the same spectral distribution.

The illuminant sources are advantageously regularly spaced around the solid support, each of said sources corresponding to a light spot, which can be automatically switched on simultaneously or successively.

The images are preferably obtained either by transparency, by reflection or by a combination thereof.

As illustrated in the enclosed drawings, the apparatus and method may comprise the use of one camera and one illuminant source, placed above the solid support, said camera and said illuminant source being movable in the three dimensions in space.

The apparatus and method may comprise also the use of two or more cameras oppositely arranged in a plane and placed above the solid support and one or more illuminant sources placed under the solid support.

5 The apparatus and method may comprise also the use of three or more cameras arranged according to a triangular plane or another regular or irregular pattern and placed above the solid support and one or more illuminant sources placed under the solid support.

10 The apparatus and method may comprise the use of one camera placed above the solid support, a first illuminant source placed above the solid support and under said camera, a second illuminant source placed under the solid support, the two illuminant sources being placed
15 almost symmetrically according to the position of the solid support.

Alternative preferred embodiments of the present invention are based upon the use of one or more illuminant source(s) and one or more camera(s) which may be
20 used according to the method of the present invention, either in combination or consecutively, said illuminant source(s) and/or said camera(s) can be maintained fixed during the lecture or can be moved according to a preferred translation or rotation movement along or around the solid
25 support comprising the specific volume of a precipitate.

It is also possible by using one or more illuminant source(s) and/or one or more camera(s) to allow the movement of the solid support comprising a specific volume of a precipitate.

30 Other embodiments that may be used according to the invention are apparatus comprising (i) either one camera and several illuminant sources with the different illuminant sources arranged from each other according to different symmetric or non-symmetric patterns, (ii) or one

illuminant source and several cameras, said cameras being arranged from each other according to different symmetric or non-symmetric patterns, (iii) or a combination thereof. The illuminant is an infra-red light having a wavelength
 5 similar to the dimension of the crystals contained in the precipitate.

The person skilled in the art is also able to provide means for performing the various steps of the present invention, especially the transformation and the
 10 conversion of the major volume into digital form or a set of digital forms by using known means or methods such as the ones present in the software and computer technologies.

The present invention is also related to a computer program product (software), comprising program
 15 code means for performing all or part of the step of the method according to the invention, when said program is run on a computer.

The present invention is related to a computer program product comprising program code means
 20 stored on a computer readable medium for performing the method according to the invention, when said program product is run on a computer.

Said means are able to collect the results obtained from said detection and/or quantification device
 25 and possibly the information(s) obtained by said reading device, and said means are able to carry out a diagnostic and/or quantification of a specific target compound resulting from the analysis of said results, possibly correlated to the read information(s).

Said means of this computer program product
 30 are able to obtain a discrimination between the spots and a possible detected background noise, for instance by the identification of homogeneous parts of an image after having been merged into two classes used as training sets.

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This discrimination can be enhanced by post-classification contextual filters techniques.

Said means are also able to identify the contour of the spot itself, which will be superposed to the original image and will allow the measure of intensity level of the counted pixels identified in the spot.

The quantification means allow an integration of all pixels intensity present in the spot or a recording the overall level of intensity of the homogeneous parts of the spot.

Furthermore, these means allow a statistical comparative analysis between the spots of each sample and a control or reference standard (standard target compound) or between two or more spots (preferably with a correlation with the recorded information of the solid support). Image correlation could be obtained between the spot image and said standard target compound spot image in order to discriminate spots that are statistically different in one test compared to another.

The recorded signal(s) by the detection device and the reading device can be read, processed as electronically computerised data, analysed by said appropriate computer program product (software).

According to a specific embodiment of the present invention, the array bears fixed oligonucleotide capture nucleotide sequences so as to allow a detection, amplification and possibility quantification of nucleic acid sequences upon a same solid support. In an alternative form of execution, the array comprises fixed PCR primers in order to obtain the production of amplicons and fixation of amplicons upon the surface according to the method described by Rasmussen et al. (Anal. Biochem., 198, pp. 138-205 (1991)), which allows thereafter their detection.

The array according to this invention is used in a diagnostic kit, in a diagnostic and/or quantification apparatus which allows automatic lecture, possibly after a previous treatment, such as purification, cleaving, copying
 5 and/or genetic amplification.

Preferably, the detection and/or quantification apparatus according to the invention is a system that combines multiple steps or substeps within an integrated system as an automatic nucleic acid diagnostic
 10 system (the steps of purification of the nucleic acid sequences in a sample, of amplification (through known genetic amplification methods), the diagnostic and possibly the quantification).

Preferred embodiments of the present invention will be described in the following non-limiting examples in reference to the figures.

Short description of the drawings

The figure 1 compares the detection of target molecules obtained on arrays composed of DNA capture nucleotide sequences covalently fixed on glass and used to detect 3 concentrations of biotinylated target DNA either in fluorescence or after silver concentration.
 20

The figures 2 to 7 represent the spatial arrangement of some elements in various embodiments of the apparatus for performing the detection and/or the quantification method according to the invention.
 25

Example 1

30 Detection of DNA on biochips

In this experiment, target DNA labelled is detected by direct hybridisation on capture nucleotide sequences bound to the array. Capture nucleotide sequences were covalently bound on glass and direct hybridisation

performed with complementary biotinylated DNA. The positive hybridisation was detected with silver precipitate catalysed by the nanogold particles linked to streptavidin.

5 Binding of capture nucleotide sequences on glass

Activated glass bearing aldehyde groups were purchased from CEL Associates (USA). Aminated capture nucleotide sequences for CMV DNA were constructed by PCR amplification of the DNA using aminated primer as described
10 by Zammattéo et al. (Anal. Biochem., 253, pp. 180-189 (1997)). The primers were purchased from Eurogentec (Liège, Belgium). Quantification of the amplicons was done by their absorption at 260 nm.

For the grafting on glass, a solution of
15 aminated amplicons at 0.2 μ m in MES 0.1 M pH 6.5 was first heated at 100 °C for 5 min and then spotted by a robot using 250 μ m diameter pins (Genetix, UK). After incubation of 1 h at 20 °C, they were washed with SDS solution at 0.1% and then two times with water. They were then incubated
20 with NaBH₄ at 2.5 mg/ml solution for 5 min then washed in water and heated at 95 °C for 3 min before being dried.

Hybridisation of the target molecule

The target molecule was obtained by
25 amplification by PCR in the presence of biotinylated dUTP at 1 mM (Alexandre et al., Biotechniques, 25, pp. 676-683 (1998)). Plasmids containing the sequence of CMV virus were used for the PCR. After amplification, the PCR products were purified using a kit of high pure PCR product
30 purification (Boehringer, Mannheim, Germany) and quantified by ethidium bromide staining after separation on a 2% agarose gel.

For the hybridisation, various concentrations 0.67, 6.7 and 67 fm in 5 μ l of biotinylated target DNA were added in a SSC 2X Denhard solution containing 20 μ g of Salmon DNA. A drop of this solution (5 μ l) was added on the array and incubated for 2 h at 65 $^{\circ}$ C in a wet atmosphere. The array was then washed 4 times with a maleic acid buffer 10 mM pH 7.5, containing NaCl 15 mM and Tween 0.1%.

10 Silver precipitation on the array after silver precipitation

The array was first incubated for 45 min with 0.8 ml of a streptavidin-colloidal gold (Sigma) diluted 1,000 times in a maleic buffer 150 mM pH 7.4 containing NaCl 100 mM and 0.1% dry milk ponder. The arrays were then washed 5 times 2 min in the maleic acid buffer 10 mM pH 7.4 containing 15 mM NaCl and Tween 0.1%. A "silver enhancement reagent" (40 μ l) from Sigma was added onto the array and changed after 10 and then 5 min. After washing in the maleic buffer, the array was dried.

20

Detection and analysis of the array

The array was scanned and the digitalised image was treated with form recognition software in order to delimitate and identify the spots. The level of the pixels of each spot was integrated and a value given to each spot. The values were corrected for the background obtained in three places where no capture nucleotide sequences have been fixed.

Example 2

Detection of proteins on biochips

Fixation of antibodies on the array

The glass of the array was activated as described here above in order to obtain aldehyde groups on the surface. The antibodies used in this experiment were raised against bovine serum albumin for positive control and non specific IgG for negative control. The antibodies at 10 $\mu\text{g/ml}$ in PBS solution were spotted using the 250 μm diameter pins directly on the glass. The amino groups of the antibodies could react with the aldehyde present on the glass. The reaction was performed for 1h at room temperature. The glasses were washed with a PBS buffer.

15 Detection of bovine serum albumin by ELISA on the array

A solution of bovine serum albumin (BSA) at 10 $\mu\text{g/ml}$ in PBS containing 0.1% casein was added on the array and incubated for 30 min. The array was then washed 3 times with PBS containing 0.1% Tween 20 and then incubated with a solution of biotinylated anti-BSA at 20 $\mu\text{g/ml}$ in PBS containing 0.1% casein. The incubation was performed for 30 min. A streptavidin-Gold complex at 1 $\mu\text{g/ml}$ was then incubated for 30 min in a PBS solution containing 0.1% casein. The presence of gold served as a center for silver reduction. The silver precipitation was performed with a "silver enhancement reagent" from Sigma with a change of the solution after 10 min and then again after 5 min. The glasses were then scanned and the data analysed as presented in the example here above.

30

Example 3

Preferred embodiments of the apparatus for performing the quantification method according to the invention is shown in the figures 2 to 7. The apparatus

comprises a solid support 1, several illuminant sources 2 regularly spaced from each other on a circular support 4, said circular support being placed under said solid support 1, and two cameras 3, 3', said cameras being placed above
5 said solid support 1 and being arranged oppositely in a plane.

The apparatus may also comprise a solid support 1, several illuminant sources 2 regularly spaced from each other on a circular support 4, said circular support being placed under said solid support 1, and one camera 3 placed above said solid support 1.

Further, the apparatus may also comprise a solid support 1. The apparatus comprises also a first set of illuminant sources 2 and a second set of illuminant sources 2', the illuminant sources of each set 2, 2' being regularly spaced from each other in a plane, preferably on a circular support 4, 4'. The first set of illuminant sources 2 is placed above the solid support 1 and the second set 2' is placed under said solid support 1, said first and said second sets of illuminant sources 2, 2' being placed symmetrically according to the position of said solid support 1. The apparatus also comprises a camera 3 placed above said solid support 1 and above the first set of illuminant sources 2.

25 Further, the apparatus may also comprise a solid support 1, with or without several illuminant sources 2 being regularly spaced from each other in a plane, preferably on a circular support 4, and being placed under the solid support 1. The apparatus comprises also a camera 30 3 placed above. Said circular support 4 and said camera 3 are placed symmetrically according to the position of the solid support 1.

Finally, the apparatus may also comprise a solid support 1 with several illuminant sources 2 regularly

spaced from each other in a plane, preferably on a circular support 4, said circular support being placed under said solid support 1, and three cameras 3, 3', 3'', said cameras being placed above said solid support 1 and being arranged according to triangular arrangement in a plane.

	1970	1971	1972	1973	1974	1975	1976	1977	1978	1979	1980	1981	1982	1983	1984	1985	1986	1987	1988	1989	1990	1991	1992	1993	1994	1995	1996	1997	1998	1999	2000	2001	2002	2003	2004	2005	2006	2007	2008	2009	2010	2011	2012	2013	2014	2015	2016	2017	2018	2019	2020	2021	2022	2023	2024	2025	2026	2027	2028	2029	2030	2031	2032	2033	2034	2035	2036	2037	2038	2039	2040	2041	2042	2043	2044	2045	2046	2047	2048	2049	2050	2051	2052	2053	2054	2055	2056	2057	2058	2059	2060	2061	2062	2063	2064	2065	2066	2067	2068	2069	2070	2071	2072	2073	2074	2075	2076	2077	2078	2079	2080	2081	2082	2083	2084	2085	2086	2087	2088	2089	2090	2091	2092	2093	2094	2095	2096	2097	2098	2099	2100	2101	2102	2103	2104	2105	2106	2107	2108	2109	2110	2111	2112	2113	2114	2115	2116	2117	2118	2119	2120	2121	2122	2123	2124	2125	2126	2127	2128	2129	2130	2131	2132	2133	2134	2135	2136	2137	2138	2139	2140	2141	2142	2143	2144	2145	2146	2147	2148	2149	2150	2151	2152	2153	2154	2155	2156	2157	2158	2159	2160	2161	2162	2163	2164	2165	2166	2167	2168	2169	2170	2171	2172	2173	2174	2175	2176	2177	2178	2179	2180	2181	2182	2183	2184	2185	2186	2187	2188	2189	2190	2191	2192	2193	2194	2195	2196	2197	2198	2199	2200	2201	2202	2203	2204	2205	2206	2207	2208	2209	2210	2211	2212	2213	2214	2215	2216	2217	2218	2219	2220	2221	2222	2223	2224	2225	2226	2227	2228	2229	2230	2231	2232	2233	2234	2235	2236	2237	2238	2239	2240	2241	2242	2243	2244	2245	2246	2247	2248	2249	2250	2251	2252	2253	2254	2255	2256	2257	2258	2259	2260	2261	2262	2263	2264	2265	2266	2267	2268	2269	2270	2271	2272	2273	2274	2275	2276	2277	2278	2279	2280	2281	2282	2283	2284	2285	2286	2287	2288	2289	2290	2291	2292	2293	2294	2295	2296	2297	2298	2299	2300	2301	2302	2303	2304	2305	2306	2307	2308	2309	2310	2311	2312	2313	2314	2315	2316	2317	2318	2319	2320	2321	2322	2323	2324	2325	2326	2327	2328	2329	2330	2331	2332	2333	2334	2335	2336	2337	2338	2339	2340	2341	2342	2343	2344	2345	2346	2347	2348	2349	2350	2351	2352	2353	2354	2355	2356	2357	2358	2359	2360	2361	2362	2363	2364	2365	2366	2367	2368	2369	2370	2371	2372	2373	2374	2375	2376	2377	2378	2379	2380	2381	2382	2383	2384	2385	2386	2387	2388	2389	2390	2391	2392	2393	2394	2395	2396	2397	2398	2399	2400	2401	2402	2403	2404	2405	2406	2407	2408	2409	2410	2411	2412	2413	2414	2415	2416	2417	2418	2419	2420	2421	2422	2
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